



Streamlining NGS Data Management & Analysis

Reads to Discovery

RNA-Seq

DNA-Seq

ChIP-Seq

Small
RNA-Seq

Methyl-Seq

MeDIP-Seq

www.strand-ngs.com

Analyze | Visualize | Annotate | Discover

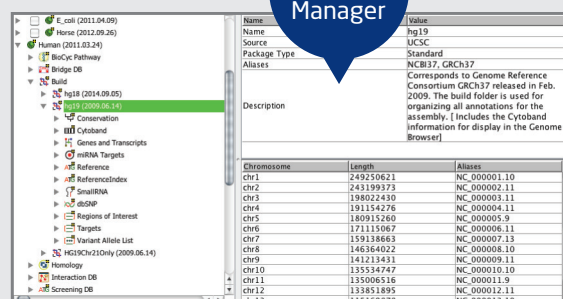
strand 
New Generation Healthcare

Data Import			Alignment	
Vendor Platforms: <ul style="list-style-type: none">• Illumina• Ion Torrent• Roche 454• ABI SOLiD• Pacific Biosciences	File Formats: <ul style="list-style-type: none">• FASTA/FASTQ• Unaligned BAM• SAM/BAM• BED, Counts data• MiSeq run folder• VCF/VAL	Library Layouts: <ul style="list-style-type: none">• Single end• Paired end• Mate paired• Directional single/paired end	<ul style="list-style-type: none">• BWT based algorithm• SSE/GPU based fast implementation• Short and long reads alignment• Single and paired end alignment• Split read alignment• Arbitrary gaps and mismatches• Multiple matches• Quality/adaptor based trimming	
Data QC				
Pre-alignment QC Plots: <ul style="list-style-type: none">• Base level quality distribution• Read level quality distribution• Base composition / quality plots		Post-alignment QC Plots: <ul style="list-style-type: none">• Alignment score distribution• Mapping quality distribution• Illumina lane/tile QC plots• Read length distribution plots• Read quality distribution plots		Filter Low Quality Reads: <ul style="list-style-type: none">• Filter by read metrics• Filter duplicate reads• Filter multiple mapping reads• Filter by tile quality• Filter by region lists
RNA-Seq	DNA-Seq	ChIP-Seq	Small RNA-Seq	
<ul style="list-style-type: none">• Gene, exon and transcript level quantification• Differential gene expression• Differential gene splicing• SNPs, MNPs and InDels• Novel genes• Novel splice junctions• Gene Fusions	<ul style="list-style-type: none">• WGS, WES, Targeted panels• SNPs, InDels and SVs• Annotate with dbSNP, COSMIC• Effect on transcripts• SIFT, Polyphen2 predictions• Multi-sample SNP analysis• Copy number analysis	<ul style="list-style-type: none">• Peak detection using PICS and MACS• TF regulation binding sites• Identify affected genes• Histone modification sites• ChIP sample vs control• Motif detection• Scan motifs in the genome	<ul style="list-style-type: none">• Quantification of miRNA, tRNA, snRNA, snoRNA and scRNA• Novel small RNA prediction• Differential gene expression• Target mRNA prediction using TargetScan, PicTar, microRNA.org, PITA	
Methyl-Seq	MeDIP-Seq	Strand NGS - Server Edition		
<ul style="list-style-type: none">• Detect hyper- and hypo-methylation• Detect DMCs and DMRs• Perform intra-sample analysis• Perform methylation effect analysis	<ul style="list-style-type: none">• Normalize using a Calibration curve• Detect hypo- and hyper-methylation• Annotate methylation call	<ul style="list-style-type: none">• Collaborative analysis• Centralized storage• Scalable compute• Web-based interface for system administration• Easy and flexible deployment		
Biological Interpretations		Discovery		
<ul style="list-style-type: none">• GO enrichment, GSEA, GSA• Single experiment OR Multi-omics analysis• Identify significant pathways• Curated / literature-derived pathway rendering• Intuitive data overlay• Create custom pathways		Rich visualizations: <ul style="list-style-type: none">• Gene view• Variant support view• Elastic genome browser	Managed Annotations: <ul style="list-style-type: none">• Pre-packaged RefSeq, UCSC, NSEMBL, and dbSNP annotations• Custom annotations• BioCyc pathways	Customization: <ul style="list-style-type: none">• Jython scripts• R-scripts• Configurable pipeline

Comprehensive Annotations

- Pre-packaged gene and transcript annotations from UCSC, RefSeq and ENSEMBL for all model organisms.
- SNP annotations from dbSNP and COSMIC, SIFT/Polyphen scores from dbNSFP, small RNA annotations, miRNA target prediction databases, screening databases and more.
- Ability to create annotations for other organisms from gbk/gtf files or FASTA files.

Annotation Manager



Chromosome	Length	Aliases
chr1	249,250,621	NC_000001.10
chr2	243,199,773	NC_000002.11
chr3	198,022,430	NC_000003.11
chr4	191,154,276	NC_000004.11
chr5	180,915,960	NC_000005.9
chr6	171,115,067	NC_000006.11
chr7	159,338,663	NC_000007.13
chr8	146,364,022	NC_000008.10
chr9	141,213,431	NC_000009.11
chr10	135,534,747	NC_000010.10
chr11	135,006,516	NC_000011.9
chr12	133,851,895	NC_000012.11
chr13	115,189,878	NC_000013.10

Import Data

- File Formats - FASTA/FASTQ, Unaligned BAM, SAM/BAM, Counts data.
- Library Layouts - Single End, Paired End, Mate Paired as well as Directional layouts.
- Vendor Platforms - Illumina, Ion Torrent, 454 Roche, SOLiD, PacBio.

Experiment Metadata

Choose Meta Data
Select the appropriate organism, build, sequencing platform and library layout.

Organism: Human
Build: Human hg19 (UCSC)
Gene Annotation: RefSeq Genes (2013.12.31)
Sequencing Platform: Illumina
Library layout: Paired End

NOTE: All reads from the sample for analysis. Make sure they are aligned to the correct reference genome.

Type	Selected files/samples	Sample Name
<input type="checkbox"/>	Leukemia-K-562-3-chr5	Leukemia-K-562-3-chr5
<input type="checkbox"/>	Leukemia-K-562-4-chr5	Leukemia-K-562-4-chr5
<input type="checkbox"/>	Melanoma-M000216-chr5	Melanoma-M000216-chr5
<input type="checkbox"/>	Melanoma-M000921-chr5	Melanoma-M000921-chr5

Buttons: Help, << Back, Next >>, Finish, Cancel

Data Files

Alignment with Strand NGS aligner

- Alignment for small RNA, DNA-Seq, ChIP-Seq and RNA-Seq data.
- Targeted region alignment for resequencing applications. Handles variable length reads, arbitrary number of gaps and mismatches and paired reads as well.
- Split read alignment for detecting long InDels and translocations.
- Options for trimming adaptors, low quality bases and screening reads against standard databases.
- Aligns ~8 million DNA reads against hg19 per hour per core on a 64GB RAM machine.

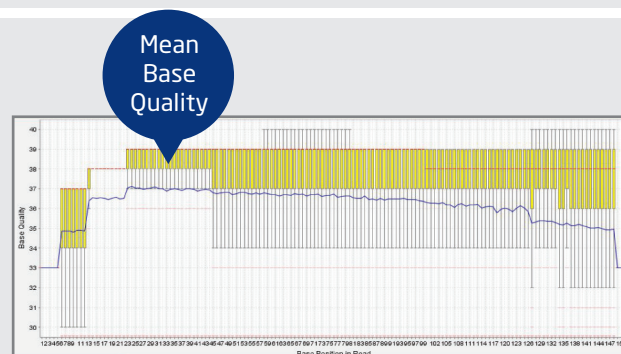
Alignment Statistics

Alignment Statistics

	H1	IM-90
Total number of reads	81,238,617 (100%)	34,881,943 (100%)
Aligned reads	73,766,623 (90.8%)	27,925,488 (80.1%)
- Uniquely matched reads	25,380,273 (72.6%)	14,830,399 (46.2%)
- Multiply matched reads	48,386,350 (59.2%)	13,095,089 (37.5%)
Unaligned reads	7,472,000 (9.2%)	6,956,455 (20.0%)
- No matches found	6,935,455 (19.9%)	6,935,455 (20.0%)
- Too many matches	0 (0%)	0 (0%)
Reads ignored due to failure of vendor QC	0 (0%)	0 (0%)
Total reads screened	81,238,617	34,881,943
Maximum read length	101	70
Average read length	36	36
Alignment Status		
Aligned to transcriptome only	21,325,609	14,946,624
- Involving known splices only	21,297,019	14,830,399
- Involving novel splices	28,590	116,225
Aligned to genome	20,941,214	13,079,864
- Involving transcriptome exons	16,975	5,075
- Without transcriptome exons	20,924,239	13,074,789
Pairs aligning to same transcript	0	0
Aligned Read Status		
Type	H1	IM-90
Single End	42,066,815	27,925,488
Unaligned	8,731,803	6,956,455
Unknown	0	0
Read Distribution		
Chromosome	H1	IM-90
chr1	4,368,888	2,421,944
chr2	5,156,266	5,888,848
chr3	1,874,154	1,093,430

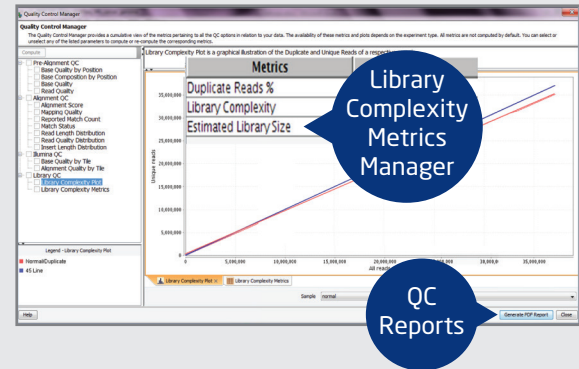
Quality Control Plots

- Base and read quality distributions.
- Base composition and quality distributions by position in read.
- Read length and insert length distributions.
- Alignment score and mapping quality distributions.
- Mate status QC plot.
- Vendor-specific QC plots.
- Targeted region QC.



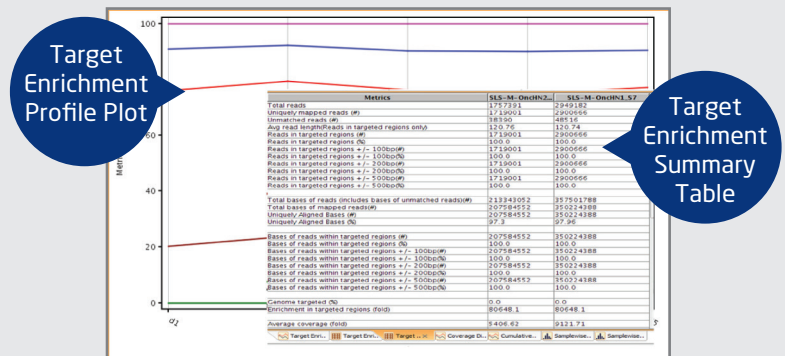
Quality Control Manager: Library QC

- Multiple quality inspection options including pre-alignment, post-alignment, vendor-specific QC and Library QC.
- Automatically generates a QC report for every sample.
- Export QC report as a pdf document.



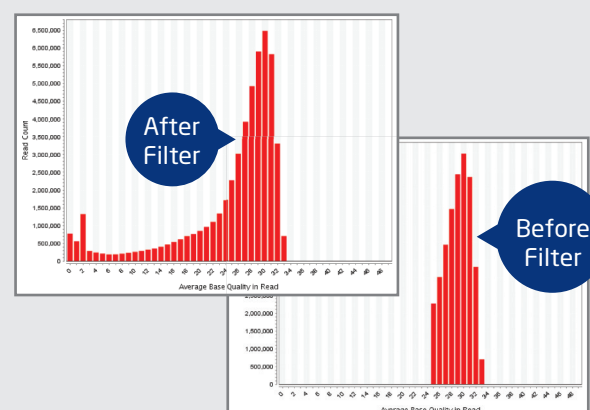
Quality Control Manager: Target Enrichment QC

- Efficiently analyze data generated from targeted resequencing experiments.
- Target region-based quality control for your customized panels.
- Assess coverage distribution across target regions including target enrichment profile plot, metrics, summary table, and sample-wise base frequency coverage distribution.

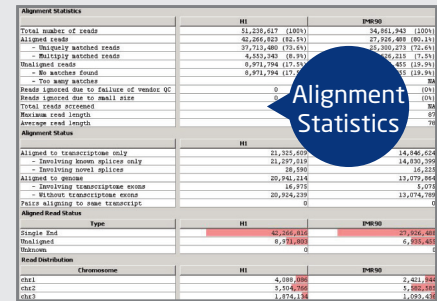


Filter Low Quality Reads

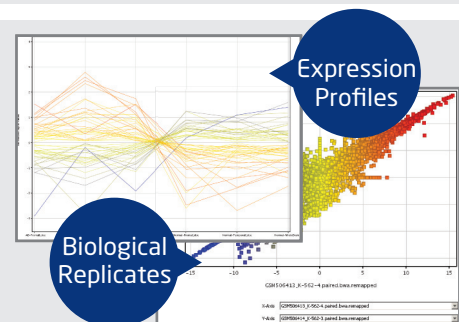
- Filter by read quality metrics.
- Filter by targeted regions of interest.
- Filter duplicate reads and multiple mapping reads.
- Filter by samples.



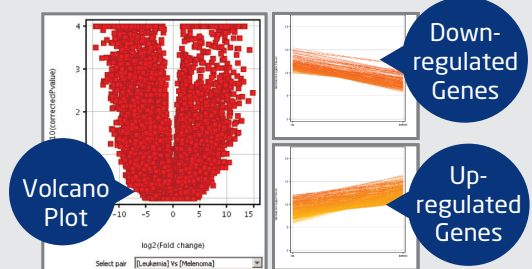
- Alignment against the transcriptome as well as the whole genome to enable detection of novel splice forms and genes.
- Handles variable length reads and paired reads as well.
- Allows arbitrary number of gaps and mismatches.
- Options for trimming adaptors, low quality bases and screening reads against standard screening databases.
- Perform RNA specific QC for genic regions and gene types.



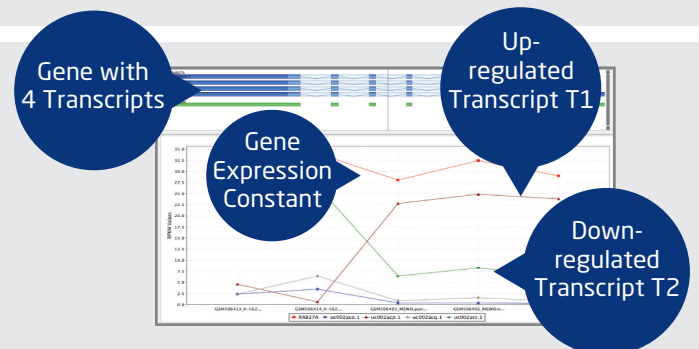
- Expression values at gene, exon and transcript levels.
- Appropriate handling of partially overlapping reads and multiple mapping reads.
- DESeq, RPM, TMM and Quantile methods for normalization.
- Experiment grouping supports large scale project handling.



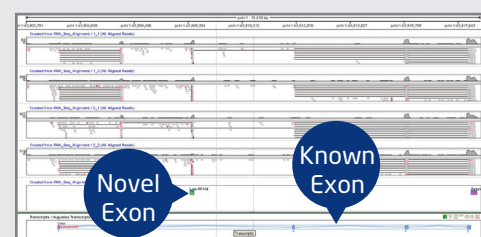
- t-Test, Mann-Whitney, N-way ANOVA for identifying differentially expressed genes under different experimental conditions.
- Multiple Testing Correction using Benjamini Hochberg, Storey, Bonferroni, etc.
- Fold Change computation and visualization.



- EM algorithm to de-convolute gene counts to transcript counts.
- Identification of differentially spliced genes.
- Visualization in the gene view.

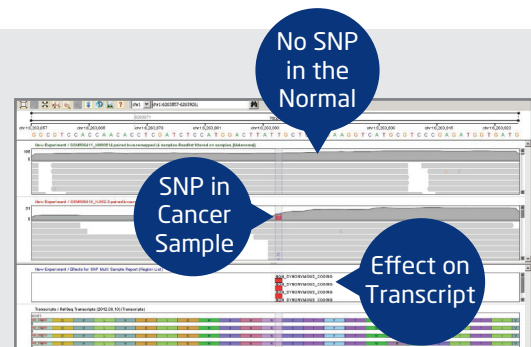


- Detection of novel genes, novel splice junctions, and novel exons in known genes.
- Differential expression analysis of novel genes and contribution of novel exons to differential splicing.
- Prioritization of novel regions using conservation score.



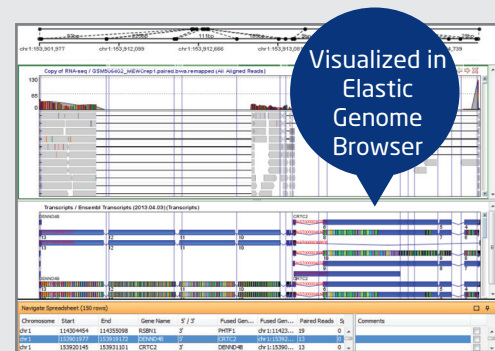
Transcriptome Variant Detection

- SNP, MNP, and InDel detection.
- Annotation with dbSNP to identify known/novel mutations.
- Prediction of effects such as non-synonymous coding, frameshift, splice-site, etc. on transcripts.
- Identify significant SNPs using an intuitive filtering framework.



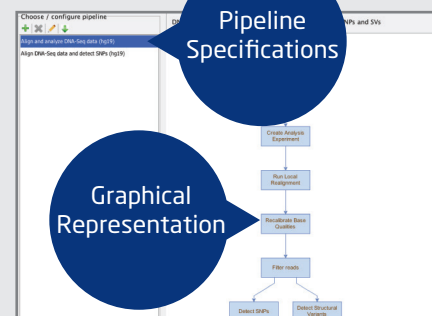
Gene Fusions

- Gene fusions detected from spliced and paired reads.
- Detection of read through transcripts.
- Gene fusions involving paralogs and pseudogenes marked as such to help filter out false positives.



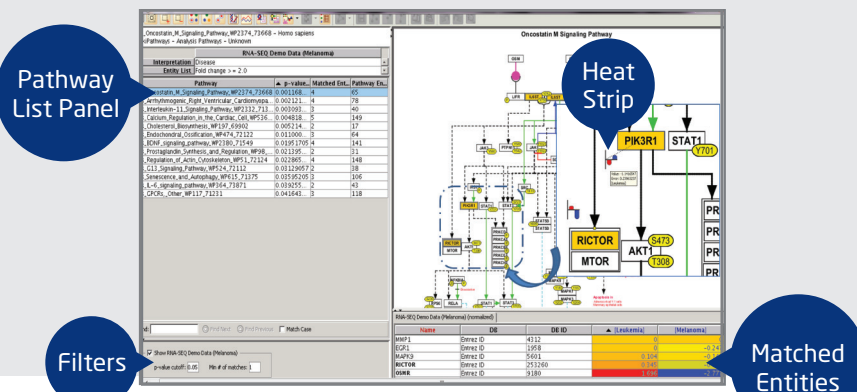
Pipeline Execution

- Pipelines that run in the background.
- Analysis pipeline includes filters and quantification.
- Pipelines support alignment of raw reads and direct import of aligned samples.
- Customization of pipelines for individual runs.

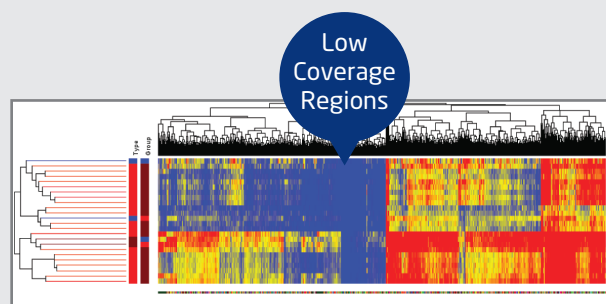


Pathway Analysis

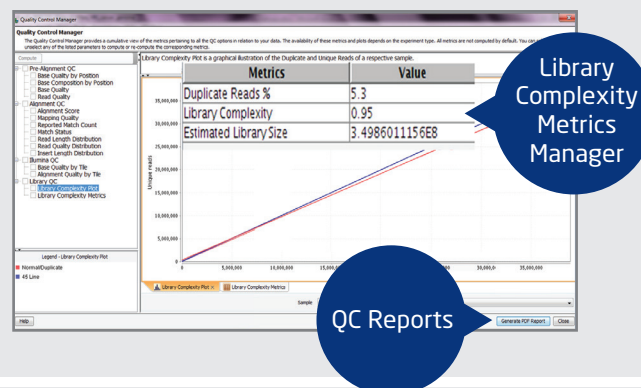
- Single Experiment Analysis (SEA) identifies enriched pathways for the genes from a single experiment type.
- Multi-Omic Pathway Analysis (MOA) from multiple genomics and transcriptomics experiments.
- Overlay differentially expressed entities on curated pathways.
- Choose from curated pathways like WikiPathways, BioCyC pathways, BioPAX pathways or literature-derived networks like NLP and MeSH.
- Find significant pathways for differentially expressed genes.



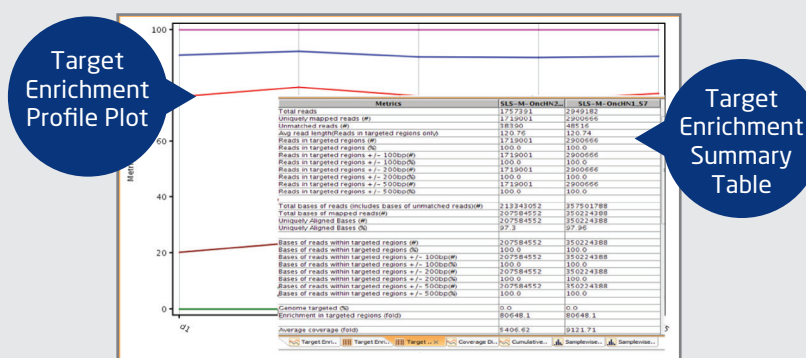
- Import BED file of target regions.
- Filter reads outside target regions.
- Evaluate efficacy of targeted re-sequencing.
- Identify target regions with low coverage across samples.
- Detect SNPs and other variants on targeted regions.



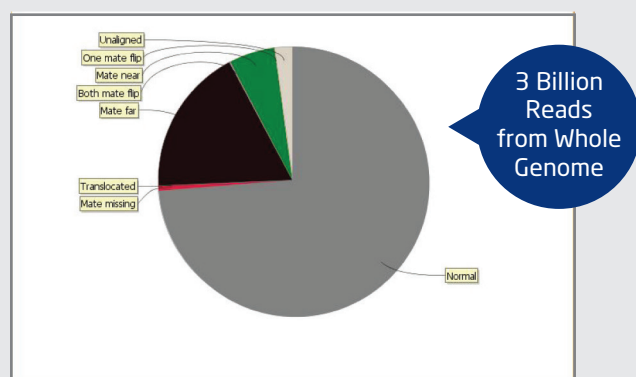
- Multiple quality inspection options including pre-alignment, post-alignment, vendor-specific QC and Library QC.
- Automatically generates a QC report for every sample.
- Export QC report as a pdf document.



- Efficiently analyze data generated from target enrichment sequencing experiments.
- Target region-based quality control for your customized panels.
- Assess coverage distribution across target regions, including target enrichment profile plot, metrics, summary table, and sample-wise base frequency coverage distribution.

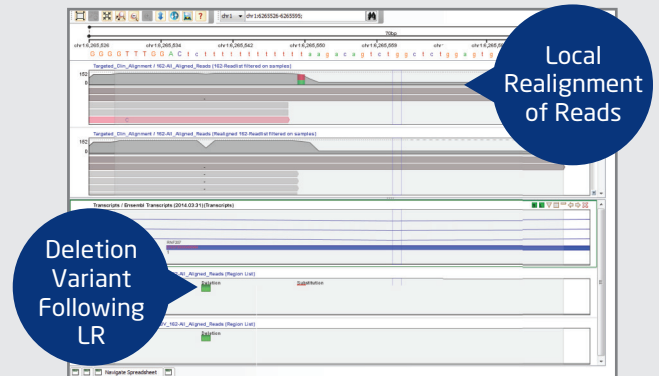


- Perform Whole Genome analysis on human or other organisms on your desktop.
- Regular desktop machine with 4GB RAM, 4 cores, and 2TB hard disk.



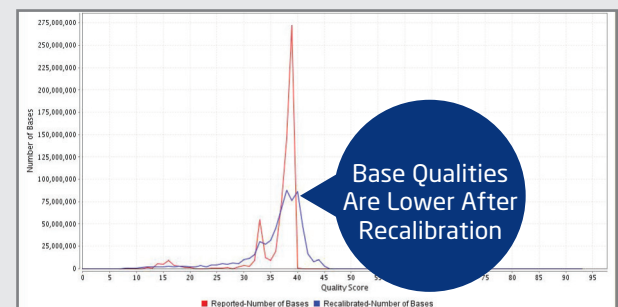
Local Realignment (LR)

- Reads with alignment artifacts around InDels can be realigned using information from multiple reads.
- Helps in getting rid of spurious substitutions and reduces false-positive variant calls.



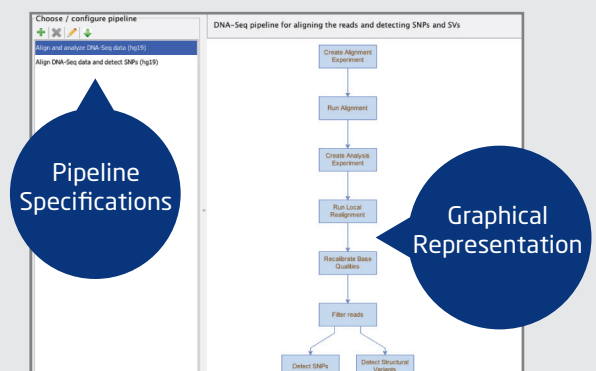
Base Quality Score Recalibration (BQSR)

- Recalibrates base quality scores to reduce errors and systemic biases.
- Uses contexts like machine cycle and dinucleotide context to recalibrate the reported base quality scores.
- Helps reduce false-positive variant calls.



Pipeline Execution

- Analysis pipelines that include filters, local realignment, base quality recalibration, and SNP detection.
- Additional pipelines that start from alignment of raw reads or direct import of aligned samples.
- Customization of pipelines for individual runs.



Variant Detection

- SNP Detection algorithm to detect SNPs, MNPs, and small InDels.
- View summary statistics of variants across samples.
- Visualize details of variants in each sample, along with dbSNP annotations.
- Support for VCF and VAL import.

Summary Statistics											
	Leukemia-K-562-3-chr5			Leukemia-K-562-4-chr5			Melanoma-M000216-c			Melanoma-M000216-c	
Substitution	300.0			302.0			346.0				
Insertion	8.0			7.0			15.0				
Deletion	12.0			7.0			15.0				
Complex	2.0			3.0			0.0				
Homozygous	129.0			113.0			111.0				
Heterozygous	190.0			203.0			264.0				
Transition mutations (Ti)	222.0			225.0			237.0				
Transversion mutations (Tv)	91.0			91.0			137.0				
TiTv ratio	2.4395604			2.4725275			1.7239271			1.94656	

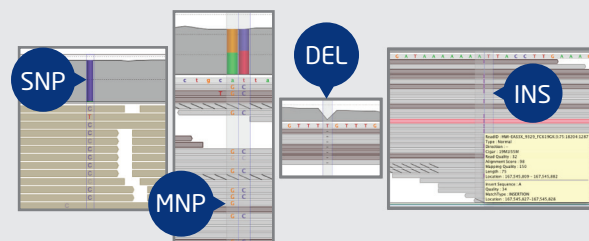
Chr	Start	End	Reference	Variant	Variant	Sample	Allele Fr.	Support	Variant	Total R.	Strand B.	SNP Call	A. Score
chr21	46444427	46444427	A	C	Substitution	1	0.083333	27.27273	36.36364	11	90.90909	A/G	50.23116
chr21	18942223	18942223	G	A	Substitution	2	0.166667	60.00000	60.00000	5	13.33333	A/G	50.24673
chr21	40198626	40198626	T	C	Substitution	4	0.333333	50.00000	50.00000	6	33.33334	C/T	52.24716
chr21	45176333	45176333	C	T	Substitution	1	0.083333	60.00000	60.00000	5	13.33333	C/T	54.23155
chr21	45180064	45180064	T	C	Substitution	1	0.083333	50.00000	50.00000	6	0.000000	C/T	54.24689
chr21	72327946	72327946	A	G	Substitution	1	0.083333	35.15152	35.15152	33	90.90909	A/G	56.93172
chr21	35515117	35515117	G	A	Substitution	4	0.500000	37.50000	37.50000	8	8.333334	A/G	58.33313
chr21	45194216	45194216	A	G	Substitution	1	0.083333	60.00000	60.00000	5	40.00000	A/G	59.27246
chr21	48069652	48069652	G	T	Substitution	1	0.083333	42.85714	42.85714	7	57.14286	G/T	61.25698
chr21	38989649	38989649	T	C	Substitution	1	0.083333	33.33334	33.33334	12	33.33333	C/T	62.09630
chr21	45140919	45140919	A	G	Substitution	2	0.166667	42.85714	42.85714	7	9.122809	A/G	62.24688
chr21	46270196	46270196	A	G	Substitution	3	0.250000	60.00000	60.00000	5	13.33333	A/G	64.26333
chr21	34636293	34636293	A	G	Substitution	1	0.083333	50.00000	50.00000	6	33.33333	A/G	64.26456
chr21	47611152	47611152	A	C	Substitution	1	0.083333	60.00000	60.00000	5	40.00000	A/C	66.26713
chr21	40670460	40670460	G	C	Substitution	4	0.333333	60.00000	60.00000	5	40.00000	G/C	68.26558

Summary on All Samples

Details of Individual Sample

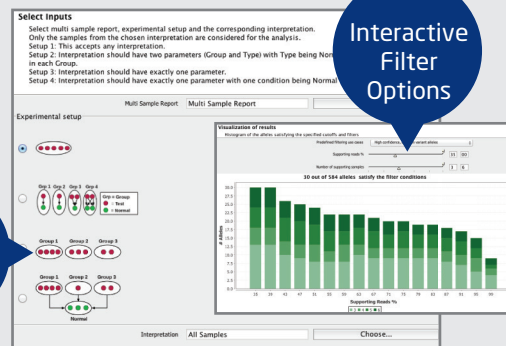
Visualize Variants

- Drag and drop SNP results into the genome browser.
- Visualize SNPs, MNPs, and InDels along with read coverage, as well as other annotations.



Find Significant SNPs

- Multiple use cases including normal-tumor, multi-group comparisons, low-frequency mutations, rare variant analysis, and somatic mutations.
- Cluster significant SNPs and samples to detect patterns visually.
- Identify significant SNPs using an intuitive filtering framework.



Multiple Use Cases

Interactive Filter Options

Variant Support View

- Intuitive visualization to verify individual SNPs.
- Color by base quality or mapping quality.
- Cluster reads to make variant locations stand out.
- Annotate clusters with strand information.

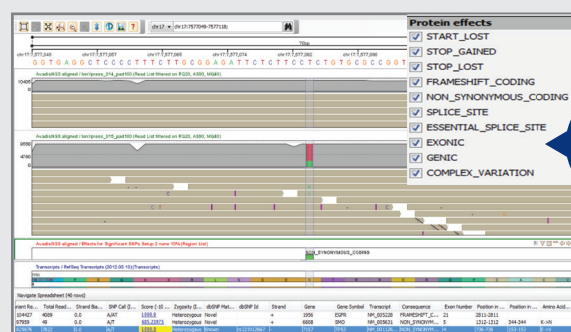
Chromo...	Start	End	Reference	Variant...	Variant...	Sample...	A
chr21	15457374	15457374	G	C	Substitution	6	
chr21	15457374	15457374	G	A	Substitution	3	
chr21	40769017	40769017	G	T	Substitution	5	0.

Cluster Id	c	t	c	a	g	c	a	g	a	c	Size
1	8
2	A	.	.	.	A	4
3	A	.	.	.	C	4
4	3
5	T	2
	
Total Coverage:	13	15	25	25	26	26	26	26	26	26	80.77 %

Clusters of Reads with Variants Marked

SNP Effect Analysis

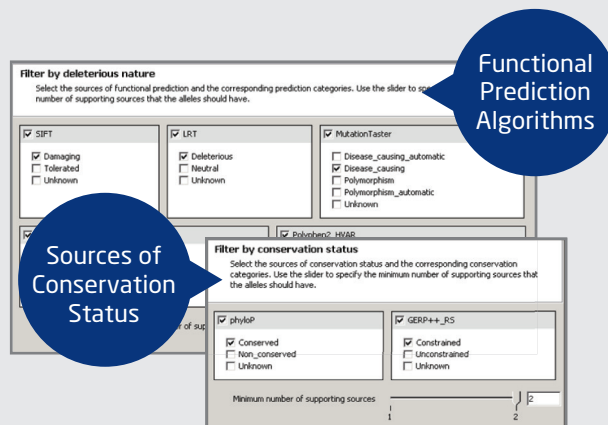
- Identify effect of detected SNPs on transcripts.
- Variety of effects including non-synonymous coding, splice site, stop gained, etc.
- Visualize along with the amino acid sequences for transcripts.
- Ability to filter on interesting effects.
- Annotate with COSMIC and other external databases.



Filter on Effects

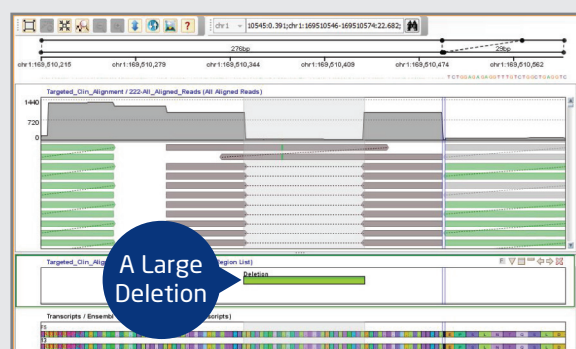
Find Damaging Variants

- Filter variants based on functional annotations for non-synonymous variants from dbNSFP.
- Identify damaging variants based on prediction scores from SIFT, Polyphen, LRT, Mutation Taster, and Mutation Assessor.
- Filter based on conservation scores from phyloP and GERP++_RS.
- Filter based on allele frequencies from 1000 genomes.



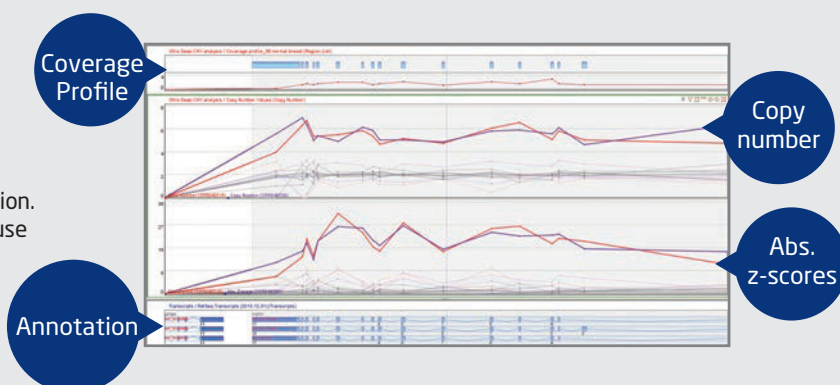
Structural Variant Analysis

- Structural variant detection on paired end and split read data.
- Identify large structural variants including large insertions, deletions, inversions, and translocations.
- Verify using Elastic Genome Browser.



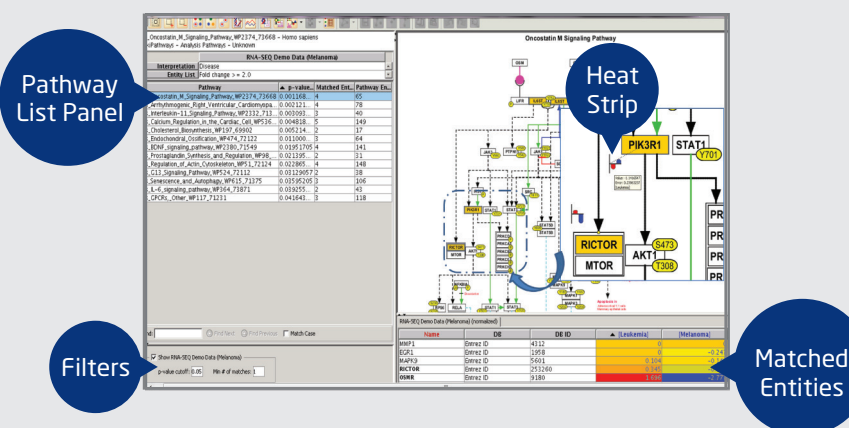
Copy Number Variant (CNV) Analysis

- Detects CNV regions in group of samples.
- Includes GC bias correction as a preprocessing step.
- Includes estimation of sample ploidy and normal cell contamination.
- Ability to create coverage profiles from a group of samples and use it for subsequent CNV analysis across experiments.
- Verify CNV's using the Genome Browser and specially designed web based visualization



Pathway Analysis

- Single Experiment Analysis (SEA) identifies enriched pathways for the genes from a single experiment type.
- Multi-Omic Pathway Analysis (MOA) from multiple Genomics and transcriptomics experiments.
- Overlay differentially expressed entities on curated pathways.
- Choose from curated pathways like WikiPathways, BioCyC pathways, BioPAX pathways or literature-derived networks like NLP and MeSH.
- Find significant pathways for differentially expressed genes.



Alignment with Strand NGS aligner

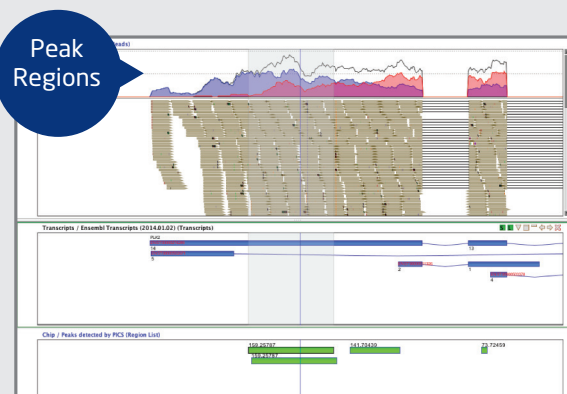
- Handles variable length reads and paired reads as well.
- Allows arbitrary number of gaps and mismatches.
- Options for trimming adaptors, low quality bases, and screening reads against standard screening databases.

Alignment Statistics			
	HI	DR90	
Total number of reads	31,236,617 (100%)	34,961,943 (100%)	
Aligned reads	42,586,823 (82.5%)	27,325,488 (80.1%)	
- Uniquely matched reads	37,713,480 (73.6%)	25,300,273 (72.6%)	
- Multiply matched reads	4,855,243 (8.9%)	2,025,215 (7.5%)	
Unaligned reads	8,971,794 (17.5%)	6,935,455 (19.9%)	
- No matches found	8,971,794 (17.5%)	6,935,455 (19.9%)	
- Too many matches	NA	NA	
Reads ignored due to failure of vendor QC	0 (0%)	0 (0%)	
Reads ignored due to small size	0 (0%)	0 (0%)	
Total reads screened	NA	NA	
Maximum read length	87	87	
Average read length	81	78	
Alignment Status			
	HI	DR90	
Aligned to transcriptome only	21,325,609	14,846,624	
- Involving known splices only	21,297,019	14,830,399	
- Involving novel splices	28,590	16,225	
Aligned to genome	20,941,214	13,979,864	
- Involving transcriptome exons	16,975	5,075	
- Without transcriptome exons	20,924,239	13,974,789	
Pairs aligning to same transcript	0	0	
Aligned Read Status			
Type	HI	DR90	
Single End	42,586,823	27,325,488	
Unaligned	8,971,794	6,935,455	
Unknown	0	0	
Read Distribution			
Chromosome	HI	DR90	
chr1	4,088,386	2,421,844	
chr2	5,504,766	5,882,585	
chr3	1,874,134	1,093,436	

Alignment Statistics

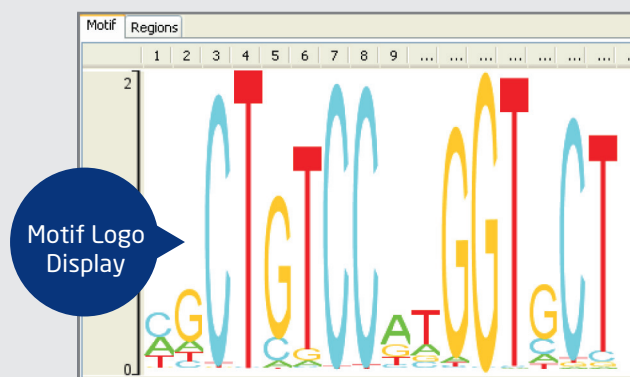
Peak Detection

- Detect enriched regions in ChIP sample as compared to the control.
- Detect peaks of transcription factor regulatory sites using PICS/MACS.
- Identify genes regulated by TF binding sites.
- Detect histone modification sites using the enriched region detection method.



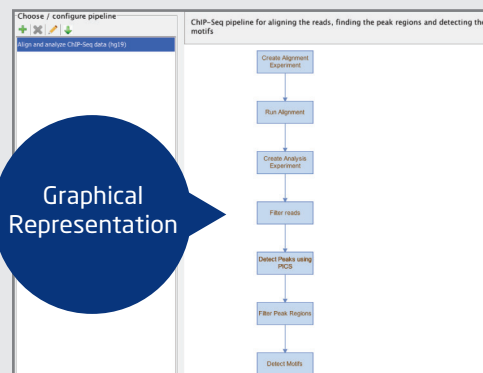
Motif Detection

- Identify motifs in detected peak regions using GADAM.
- Import peak regions to detect motifs.
- Import motifs in JASPAR format.
- Scan for motifs in the entire genome or in regions of interest.



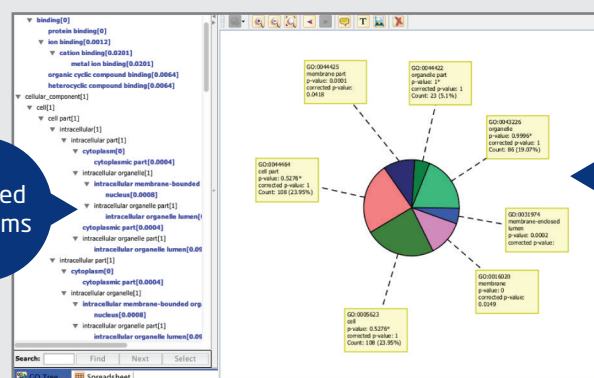
Pipeline Execution

- Pipelines that run in the background.
- Analysis pipeline that includes filters and peak detection.
- Additional pipelines that start from alignment of raw reads or direct import of aligned samples.
- Customization of pipelines for individual runs.



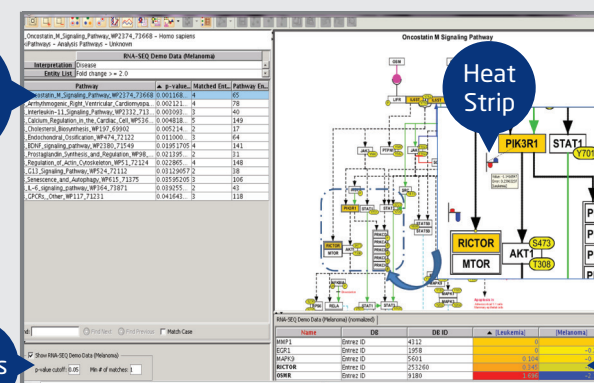
GO Enrichment

- Annotate regions with genic information to discover those affected by peak regions.
- GO Enrichment analysis to detect enriched Gene Ontology terms.



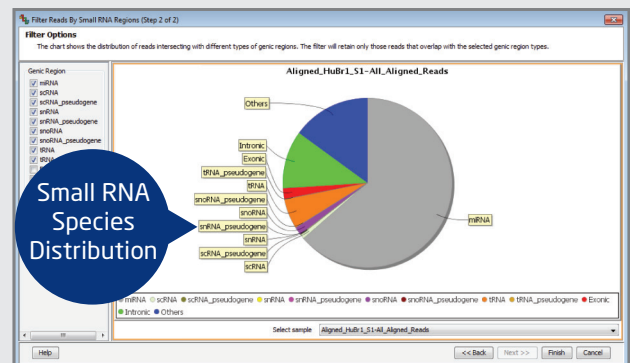
Pathway Analysis

- Single Experiment Analysis (SEA) identifies enriched pathways for the genes from a single experiment type.
- Multi-Omic Pathway Analysis (MOA) from multiple genomics and transcriptomics experiments.
- Overlay differentially expressed entities on curated pathways.
- Choose from curated pathways like WikiPathways, BioCyC pathways, BioPAX pathways or literature-derived networks like NLP and MeSH.
- Find significant pathways for differentially expressed genes.



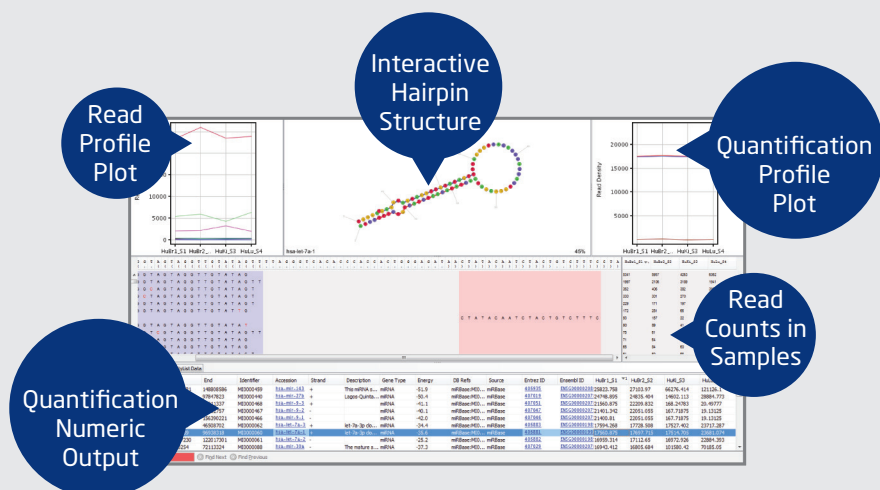
Alignment, QC and Filtering

- Read alignment with Strand NGS aligner.
- Option for trimming adapters, low quality bases and screening reads against standard screening databases
- Read distribution across genic regions and small RNA species.



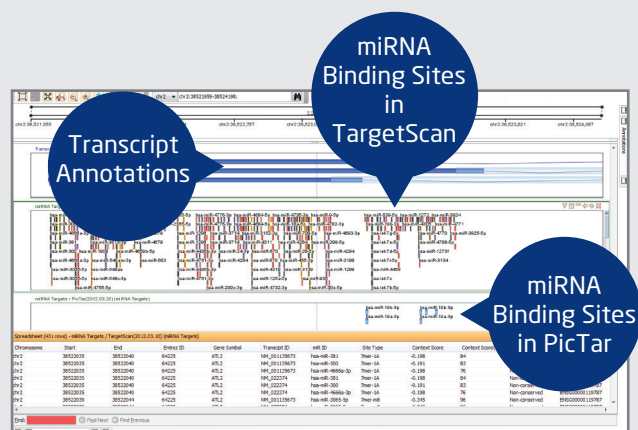
Quantification & Normalization

- Expression values for known genes, novel genes, mature miRNAs.
- Ability to pick reads aligning exactly with the 5' end of mature miRNAs.
- Take into account padding and multiply mapping reads.
- DESeq, TMM, Quantile, and Sample Count based methods for normalization.
- Small RNA Gene View to visualize quantification results.



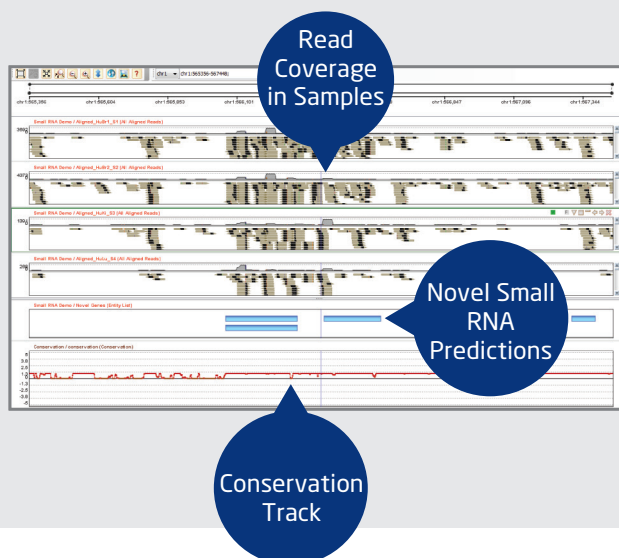
miRNA Target Analysis

- TargetScan, PicTar, TarBase, microRNA.org, and PITA databases for target prediction analysis.
- Identify targets common to multiple databases.
- Perform downstream analyses (GO, GSA, GSEA, Pathway Analysis) on target set of mRNA genes.



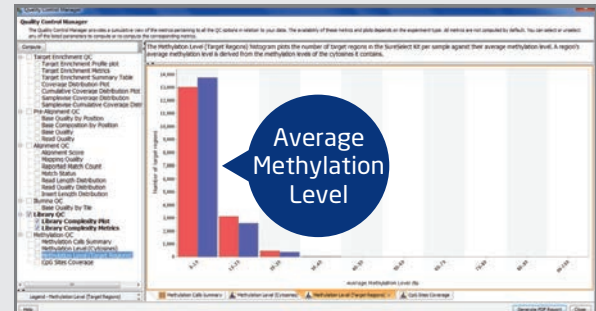
Novel small RNA Discovery

- Predict type of novel gene as one of miRNA, snoRNA, scRNA or tRNA.
- Identify high-confidence predictions with Confidence values and Conservation scores.
- Find annotation discrepancies of known genes from the read coverage patterns.



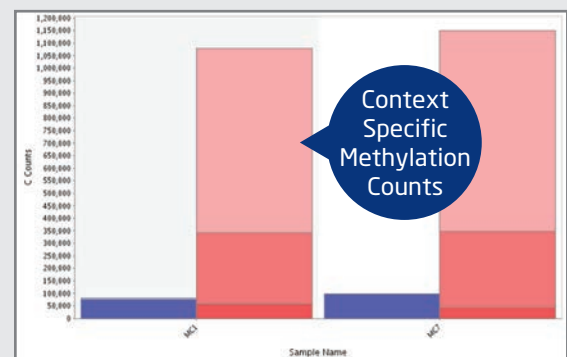
Data Import and QC

- Allows import of Bismark aligned bisulfite treated sequencing data.
- Quality metrics for pre- and post-alignment, target enrichment and library complexity.
- Compute methylation-specific QC to get information on methylation levels and CpG sites coverage for your samples.



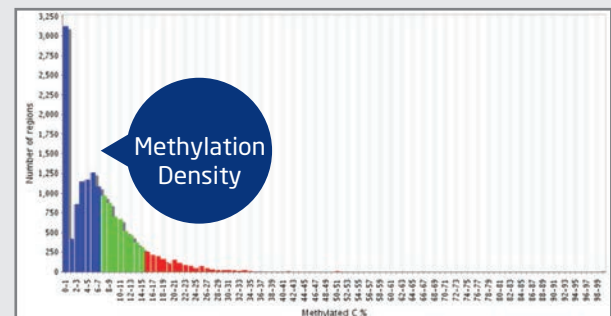
Methylation Detection

- Identify methylated cytosines for specific loci on the genome for individual samples.
- Algorithm considers bisulfite conversion error rate, sequencing error, read coverage, base quality and methylation fraction.



Intra Sample Analysis

- Identify regions with low, moderate, and high methylation density within an individual sample for target regions of interest.



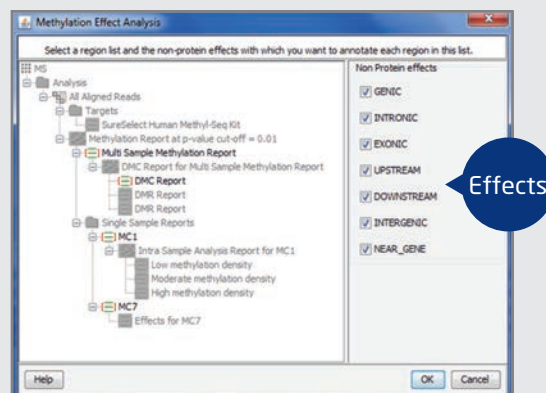
Differential Methylation

- Identify differentially methylated cytosines (DMCs) across experimental conditions/samples.
- Discover differentially methylated regions (DMRs) across experimental conditions/samples.



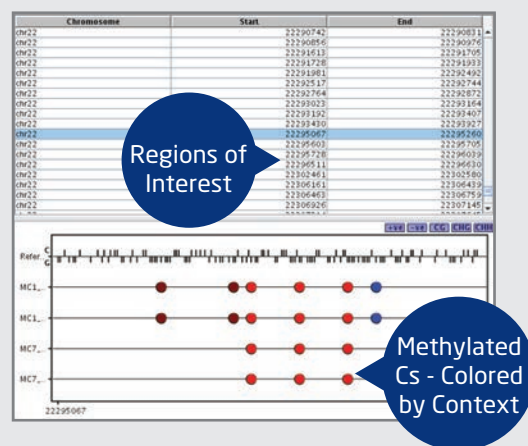
Methylation Effect Analysis

- Determine the Genomic context of the methylated or differentially methylated cytosines resulting from methylation detection and DMC analyses.
- Annotate these cytosines with the selected non-protein effects.



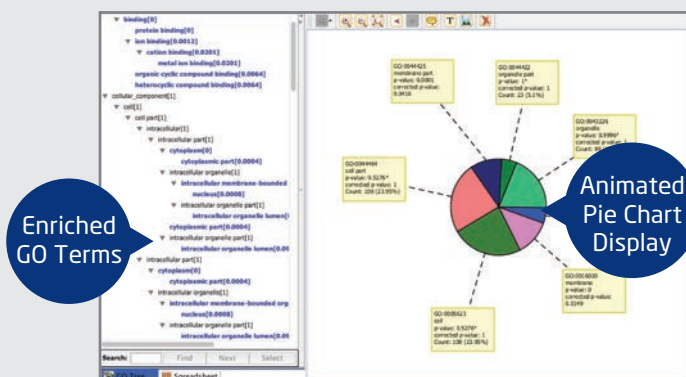
Methylation-Specific Views

- Lollipop Plot - Visualize methylated or differentially methylated cytosines by regions and samples.
- Interactive genome browser - Customized for displaying bisulfite converted reads.
- Within the GB, the methylation level histogram helps visualize the proportion of methylated cytosines compared to unmethylated cytosines in the read coverage.



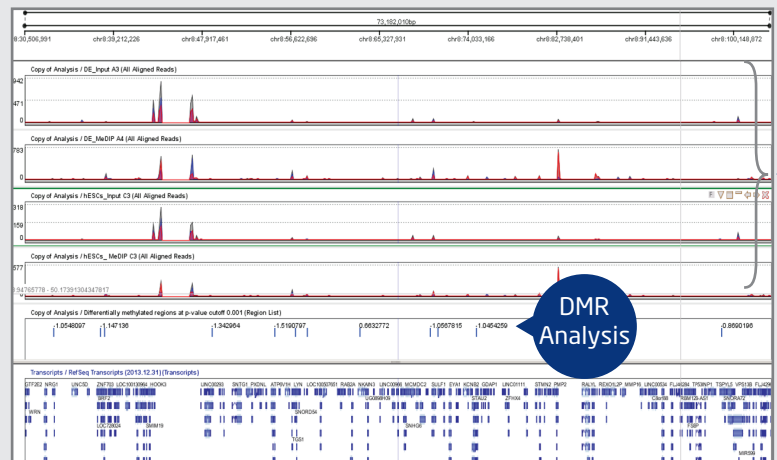
GO Enrichment

- Genes discovered to be affected by SNPs, SVs, peak regions, or any imported set of genes.
- GO Enrichment analysis on gene lists to detect enriched Gene Ontology terms.



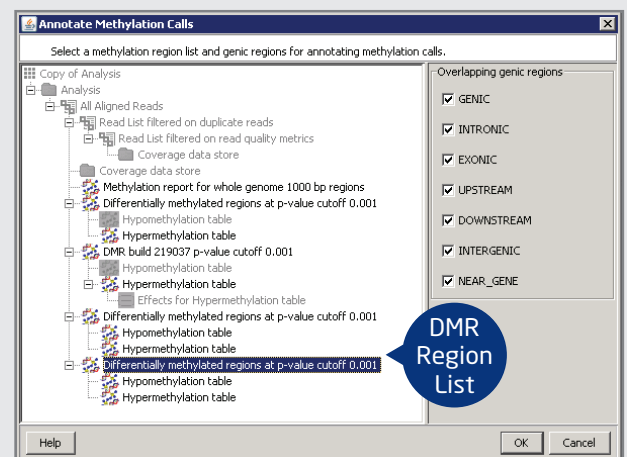
Differential methylation analysis (DMR)

- Identifies DMRs based on selected RMS ratio and p-value thresholds.
- Merges consecutive hypo and hyper-methylated regions if the new region still satisfies the specified threshold.
- Filters DMRs based on RPM, RMS or AMS and p-value thresholds.
- Creates separate lists for hypermethylated and hypomethylated regions from the identified DMRs.
- Creates associated gene lists based on an upstream / downstream padding (bp) selected for the genes.



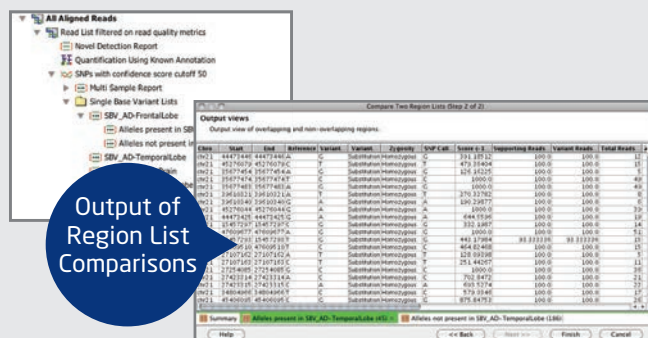
Annotate methylation calls

- Determines the genomic context of the methylated or differentially methylated regions.
- Annotates the methylation calls for the selected genic region.



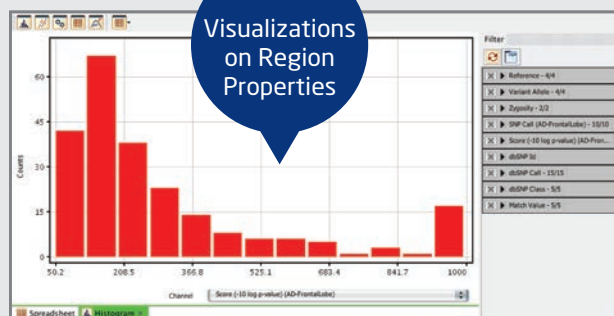
Compare Region Lists

- Choose two region lists to compare.
- Find overlapping and non-overlapping region with appropriate distance criteria.



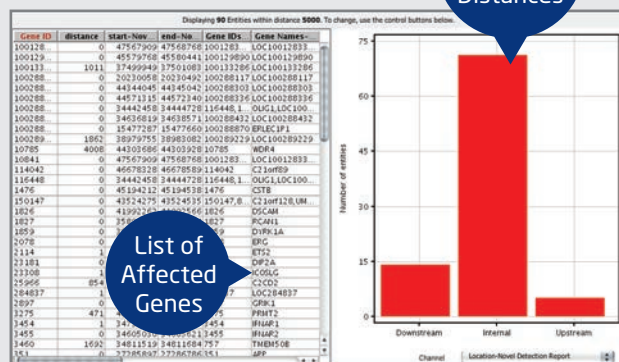
Region List Operations

- Visualize plots on various columns of a region list.
- Create new columns from existing columns using formulas.
- Perform filter operations on the columns to retain relevant regions.



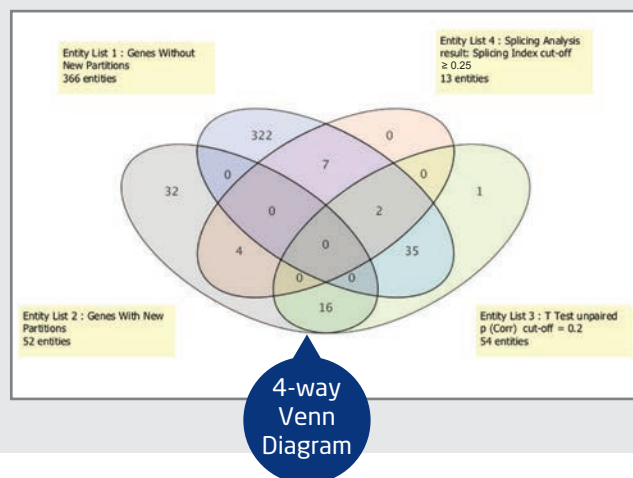
Translate from regions to genes

- For any region list find adjacent or overlapping genes.
- Find affected genes for the detected regions (eg. SNPs, SVs, peaks).



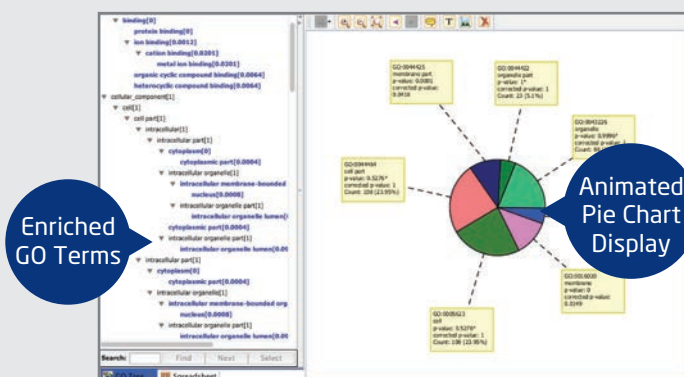
Compare Gene Lists

- Venn Diagrams to compare gene lists.
- Compare gene lists from different experiments and organisms.
- Ability to compare imported gene lists.
- Ability to save individual regions as new gene lists.



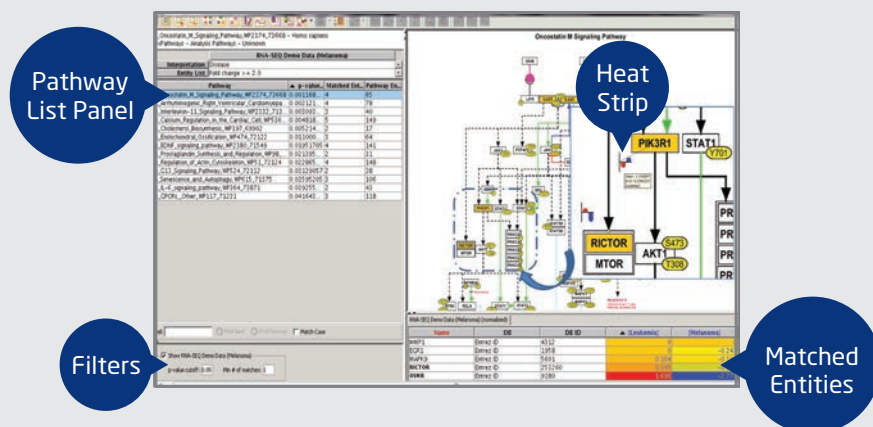
GO Enrichment

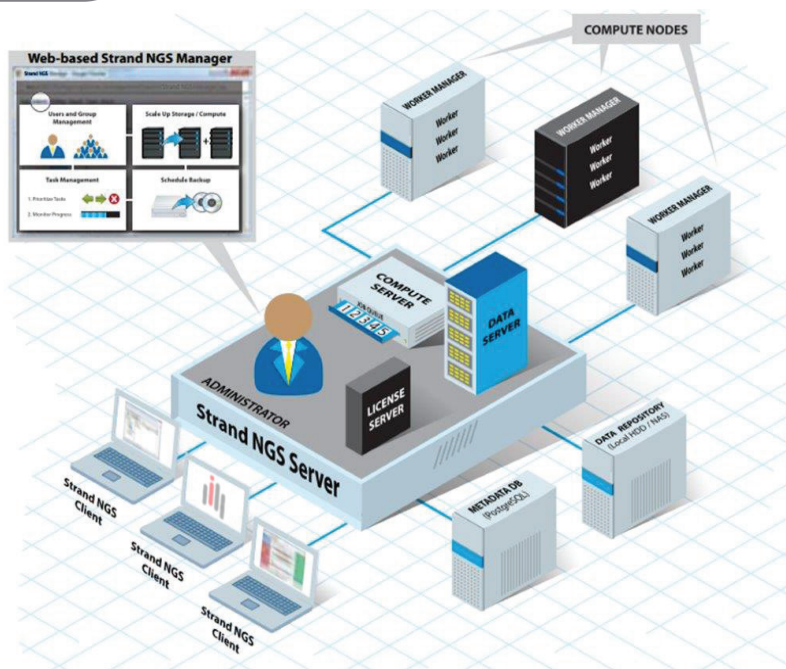
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Pathway Analysis

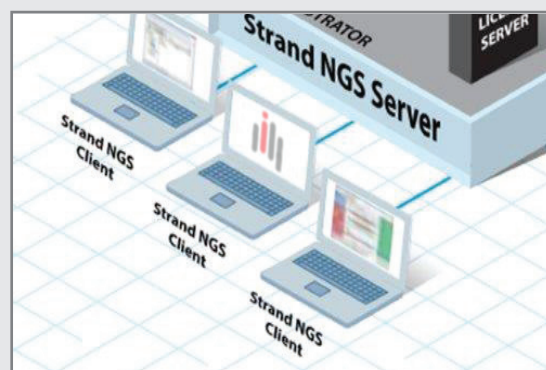
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- Choose from curated pathways like WikiPathways, BioCyC pathways, BioPAX pathways or literature-derived networks like NLP and MeSH.
- Find significant pathways for differentially expressed genes.





Collaborative Analysis

- Allows different people working on the same project to easily collaborate with each other.
- Samples, analysis, and workflow pipelines can be shared.
- Objects can be shared with an individual user or a group.
- Sharing can be done with either Read-only or Read-Write permission.



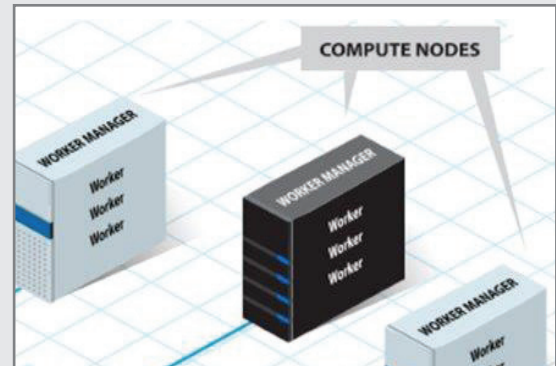
Centralized Storage

- Data and analysis results from multiple users stored in one place in a safe, backed-up manner.
- Data can be backed up regularly; possible to repeat backup weekly or monthly.
- Option to do the backup incrementally.
- Allows storing data in a new location, in case the current location is full, making incremental expansion of storage possible.



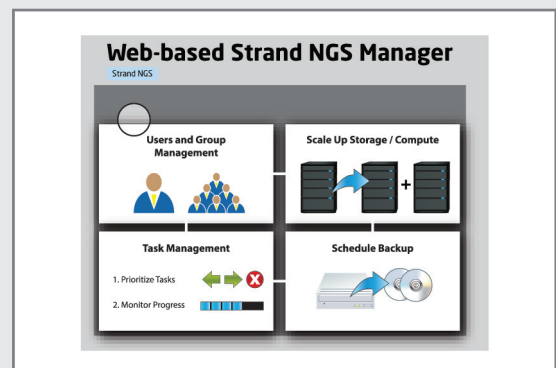
Scalable Compute

- Multiple compute nodes managed by the server to handle heavy computing.
- Compute can be scaled up by simply adding new machines to the network and configuring them as compute nodes.
- Higher reliability due to the distributed nature of the compute.
- Execution of tasks on a first-come, first-serve basis with a possibility for the administrator to reorder them if necessary.
- Status dashboard to show the task status and logs.



Web-based Interface for System Administration

- Allows creating and managing users and groups.
- Facilitates managing the compute resources.
- Allows the administrator to suspend, reschedule, reorder, or delete tasks.
- Group administrator sees only the tasks from his group members.
- System logs and usage can be monitored.



Easy and Flexible Deployment

- Major components of the Server Edition have separate installers and can be installed independently.
- Possible to have the Server, Compute, and the Storage on one big machine or on different machines depending on the available resources and the needs of the enterprise.
- Can have either node-locked licenses or shared floating licenses depending on the need.

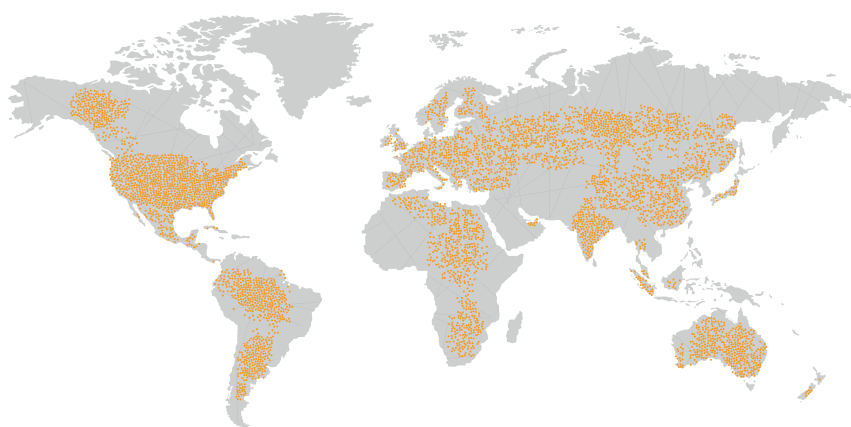


About Strand

A History of Innovative Genomic Research

Strand Life Sciences is a global genomic profiling company and leader in precision medicine diagnostics, aimed at empowering cancer care and genetic testing for inherited diseases. Strand works with physicians and hospitals to enable faster clinical decision support for accurate molecular diagnosis, prognosis, therapy recommendations, and clinical trials. The Strand Center for Genomics & Personalized Medicine is India's 1st and only CAP & NABL accredited NGS laboratory.

www.strandls.com



A Trusted Partner to Companies Worldwide

For 15 years, our genomics products and solutions have facilitated the work of leading researchers and medical geneticists in over 2,000 laboratories and 100 hospitals around the world.

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